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Received: 13 February, 2018

Accepted: 26 February, 2018

Published: 27 February, 2018

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Keywords: Broiler chicken; Gut; Morpho-histology; Phytogetic products

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Short Communication

Effects of phytogetic products on gut morpho-histology of broiler chickens

Abstract

The objective of this experimental survey is to distinguish the effect of phytogetic products (Volarom®) as an alternative to antibiotic growth promoters (AGP) on the gut morpho-histology of broilers chickens, reared under optimal environmental conditions. Undred 1-day- Cobb strain chick males were divided into 2 groups (control and experimental). The broiler chickens received the following feed: control group: basic feed, experimental group: basic feed + 50 ml of phytogetic complex / 50 chicks in water for 42 days. At the age of 42 days, five birds per treatment were randomly selected and slaughtered. Representative segments of the gut broiler chickens (duodenum, jejunum and ileum) were removed from each slaughtered bird and then subjected to fixation and histological observation. Ten consecutive villi were analyzed, measuring their height and depth. Height was measured from the base of the villi at the junction of the crypt to the tip of the villi. For morphometry the software "Image J1-45s" was used Version 1.6.0_20. The gut morpho-histological structure of broiler chickens results indicated that the addition of 50 ml of phytogetic products / 50 chicks in water for 42 days decreased respectively the villi height of the duodenum, jejunum and ileum (1230 $\mu\text{m} \pm 270$, 1127 $\mu\text{m} \pm 290$, 920 $\mu\text{m} \pm 220$). However for the depth of crypts, the phytogetic effect is found only at the level of the duodenum (238 $\mu\text{m} \pm 50$ vs 245 $\mu\text{m} \pm 60$).

Statistically, there was no effect of the phytogetic products on the different parts of the broilers gut, in the experimental group at 42 days.

Introduction

Feed additives are products used in animal nutrition to improve the quality of food animal origin, or to increase the performance of production and health of animals [1]. However, the exaggerated use AGP has resulted in increased antibiotic residues in animal products and the development of antimicrobial resistance. This led, the European Union to ban the use of antibiotics growth promoters (AGP) in livestock production, in 2006. This has resulted in the search for alternatives to AGP in animal feeds, which will provide the same protection against pathogenic and non-pathogenic microorganisms, as well as improving the performance of animals. An intensive search for substitutes such as probiotics, prebiotics, symbiotic, enzymes, organic acids, organic minerals, oligosaccharides and other feed additives has taken place over the last decade [2]. Among the potential candidates, phytogetic represents a new and exciting group of feed additives, mainly from herbs, spices or other plants. Plant extracts and essential oils have received a great deal of attention because of their natural antimicrobial properties [3], to improve immune property [4], the ability to manipulate microorganism intestinals [1]; Also they have coccidiostatic activity [5,6]. This study tested the effect of the phytogetic complex (Volarom®) on the gut morpho-histology of broilers chickens at 42 days.

Material and Methods

Description of the experimental protocol

This study was conducted at the Experimental Poultry Station, Veterinary Department, Batna1 University, during September and October 2016. Animals are raised under optimized environmental conditions (temperature, humidity, air speed, ventilation). Experimental procedures followed the principles of the Animal Care Committee of the CRBt (National Center for Biotechnology Research), Constantine, Algeria. A total of 100 broiler chickens "Cobb", male, aged 1day, were randomly allocated to 2 dietary treatments: one based on no feed additives (control), and one supplemented with 50 ml phytogetic products (Volarom®)/50 subjects in water (experimental). Feed formulation was carried out according to NRC (1996) [7], recommendations (Table 1). This phytogetic product is composed of a mixture of thyme, olive leaf, rosemary, grapefruit pepins and turmeric. Five birds per treatment were randomly selected and slaughtered, representative segments of the small intestine (duodenum, jejunum, and ileum) were taken from each bird and then subjected to fixation and histological segmentation as described by the method of [8]. Ten consecutive villi were analyzed, measuring their height

and depth [9]. For morphometry the software “Image J 1,45s” was used Version 1.6.0_20. The images are taken thanks to the use of a camera integrated into the microscope (Zeiss-Axioskope 20). The sensitivity of this camera is set to that of the film used (100 ASA).

Statistical analysis

All data were analyzed with Stat Soft software (Statistica 7, 2005). The statistically significant differences in treatment means were evaluated using one-way ANOVA, and a multiple comparison (Duncan) test was conducted with significant treatment means. $P < 0.05$ was considered as the significant level. Values are expressed as the means \pm standard deviation (SD).

Results and Discussion

The results of gut morpho-histology for broilers slaughtered at the age of 42 days are shown in table 2, indicating villous height and crypt depth. Significant differences between dietary treatments (control and experimental) were measured for duodenal villus height and crypt depth (Figure 1). Villous height was longer for broiler chickens in the control group and shorter for broilers supplemented with the phytochemical products (PP). The depths of the duodenal crypt were significantly deeper in animals supplemented with PP, and shallower for broilers in the control group.

The size of the jejunum villi was longer in broiler chickens in the control group, while the broiler chickens receiving PP had the shortest jejunum villi. The depths of the jejunum crypt were shorter in broiler chickens supplemented with phytochemical products (PP) and deeper in broiler chickens in the control group (Figure 2). Statistically, there was no significant difference between villous size and crypt depth between control and experimental groups ($p < 0.05$).

Table 2: Gut morphohistology of broilers chickens (42 days) \pm standard deviation (SD).

Gut Groups	Duodenum (n=5)		Jejunum (n=5)		Ileum (n=5)	
	Villus height of crypts (μm)	Depth of crypts (μm)	Villus height of crypts (μm)	Depth of crypts (μm)	Villus height of crypts (μm)	Depth of crypts (μm)
Control (basic diet)	1320 \pm 310	238 \pm 50	1131 \pm 100	230 \pm 70	1050 \pm 140	170 \pm 50
Experimental (+PP)	1230 \pm 270	245 \pm 60	1127 \pm 290	210 \pm 60	980 \pm 220	130 \pm 40
P*	0.001	0.1	0.7	0.07	0.07	0.0009

P *: Averages within the same column are not significantly different ($P < 0.05$); PP: Phytochemical products.

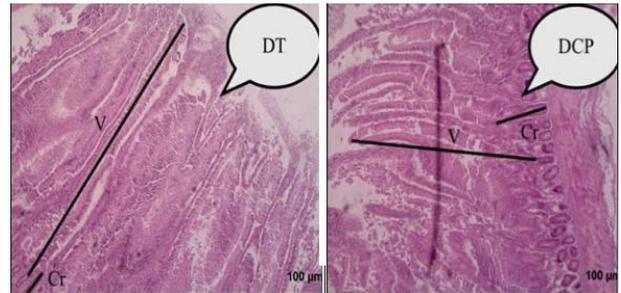


Figure 1: Histological structure of duodenum of broilers at 42 days fed with different experimental treatments. DT: control duodenum, DCP: duodenum of the phytochemical products, V: villosity and Cr: Lieberkühn crypt. (H & E - 10X).

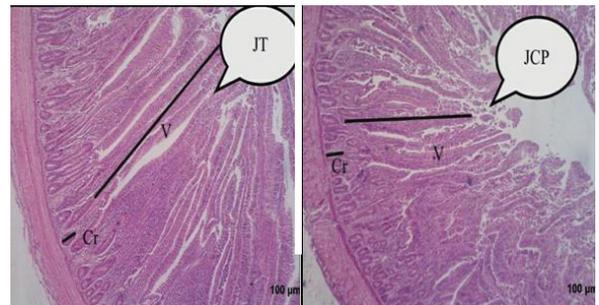


Figure 2: Histological structure of jejunum of broilers at 42 days fed with different experimental treatments. JT: control jejunum, JCP: jejunum of the phytochemical products, V: villosity and Cr: Lieberkühn crypt. (H & E - 10X).

Table 1: Composition of the basal diets of broilers chickens.

Ingredients (%)	Starter (1-21 day)	Finisher (22-42 day)
Maize	51	53
Soybean meal	30	25
Wheat	13.8	15
Oil	1	25
Premix	2.5	2.5
Di-Calcium phosphate	1.2	1.2
Salt	0.3	0.3
DL Methionine	0.1	0.1
L-Lysine	01	01
Calculated chemical analysis		
ME (Kcal/kg)	3000	3085
Crude protein %	21.2	19.4
Avialable phosphore %	0.73	0.68
Calcium %	0.70	0.53
Methionine %	0.34	0.30
Lysine %	1.19	1.08

The results of the morphohistology of the ileum show that the broilers in the control group had the highest height and depth of the crypt compared to the experimental group (Figure 3). However, the difference between the 2 groups is only significant for the depth of crypts values ($p < 0.05$).

Similar results have been found in studies by [10,11]. Recent study, revealed a significant increase in villous height, depth of crypt in broiler chickens supplemented with garlic powder at 0.5 g / kg or the combination garlic and black seed with a rate of 0.5 g / kg combined [12]. Improved intestinal morphology broilers, having consumed a ration supplemented with garlic powder or a combination of garlic powder and black seed also experienced a significant improvement in production performances [12].

Longer villi with more enterocyte cells for uptake and deeper crypts showing greater demand for new tissues due

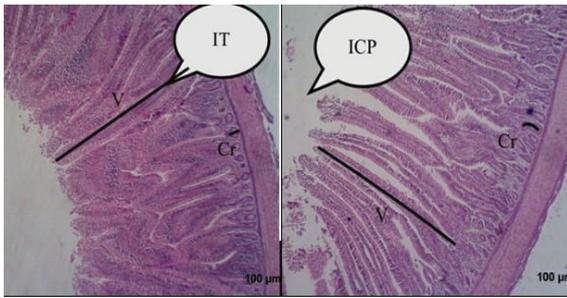


Figure 3: Histological structure of the ileum of broilers at 42 days fed with different experimental treatments. IT: control ileum, ICP: ileum of the phyto-genic products, V: villosity and Cr: lieberkühn crypt. (H & E - 10X).

to proliferation in light have been found by [13,14]. A higher turnover of cells, thus a deeper crypt, would lead to a higher energy demand for maintenance of the digestive tract of the broiler. The energy could instead be used for growth and improved performance of animals [15]. Pathogenic bacteria present in the gastrointestinal tract are considered to damage enterocyte cells, which reduces their absorption, as well as deeper crypts [13]. Phyto-genic feed additives are thought to reduce the number of pathogenic bacteria in the gastrointestinal tract [1,16], and would result in less damage to enterocytes and better uptake and villous height. However, many studies show no effect of phyto-genic products on gut morphology and histology, making it difficult to link improved intestinal morphology to better animal performances.

Conclusion

The phyto-genic product used in this experiment has an effect on the morpho-histological structure of the chicken gut (crypts and villi). However, the comparison of the results obtained between the experimental and the control group is not always statistically significant.

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