Diagnostic accuracy of skin prick test and specific IgE and their association with Total IgE, AEC and serum cortisol in Indian patients with respiratory allergy

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Abstract

Background: Indian subcontinent has high burden of patients suffering from respiratory allergy associated with food sensitization. Immune response is different in skin and blood for allergen sensitization. Skin Prick Test (SPT) and Specific IgE (sIgE) allergen positivity is an evidence of sensitization. Both these tests (SPT & sIgE) are often used interchangeably in clinical practice of allergic disorders. In our study the concordance relevance of various tests SPT, Total IgE, Absolute Eosinophil Count (AEC) & Serum Cortisol was investigated in 75 patients of respiratory allergy. The aeroallergens were divided into four groups (Dust mite, Pollen, Fungi, Cockroach) & 7 Foods (Milk, Soybean, Wheat, Fish, Peanut, Egg white, Yeast). Total 35 common allergens were tested by SPT. Serum Specific IgE estimation was carried out with 16 allergens (D. pteronyssinus, D. farinae, Alternaria alternata, Candida albicans, Aspergillus fumigatus, mesquite (Prosopis juliflora), common pigweed, goosefoot lamb’s quarter (Chenopodium album), cockroach and seven food allergens (milk, soybean, wheat, fish, peanut, egg white, yeast).

Methods and findings: A retrospective stratified sample of 75 respiratory allergic patients within age group of 05-70 years was included in the study. Serum total IgE and specific IgE levels were estimated by using ImmunoCAP® system and SPT was done by standardized allergens (Allergo Pharma and Greer laboratories Inc.).

Following results were compared to evaluate concordance/ discordance between various markers for respiratory allergy.

1. Sensitizations pattern based on Serum (16 specific IgE >1 kU/ml) with Skin prick tests Positive >4 mm wheal Size (35 aero-allergens)
2. Serum Total IgE levels >250 IU/ml with Sensitizations based on Serum (16 specific IgE >1 kU/ml) with Skin Prick Tests Positive >4 mm wheal Size (35 aero-allergens)
3. Absolute Eosinophil Count >200 cells/uL with Sensitizations based on Serum (16 specific IgE >1 kU/ml) with Skin Prick Tests Positive >4 mm wheal Size (35 aero-allergens)
4. Serum cortisol level <10 mcg/dL with Sensitizations based on Serum (16 specific IgE >1 kU/ml) with Skin Prick Test Positive >4 mm wheal Size (35 aero-allergens).

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Introduction

There is a consistent increase in allergic diseases from 10 to 30% for the last 50 years in India [1]. The global prevalence of Allergic Rhinitis is between 10 to 30% for adults and as high as 40% for children [2]. SPT and Measurement of sIgE are important tools for diagnosing atopic sensitization and both tests are often used interchangeably. There is no universally acceptable gold standard cut off value for either of these tests associated with allergic diseases, although practice parameter third update guidelines recommends that wheal size threshold of >3mm cutoff criteria has better clinical accuracy which is based on reproducibility in relation to nasal provocation test. There are several factors which can modify the test such as skill of the tester, the testing device, the color of the skin, skin reactivity on the day of testing, potency and stability of allergen extract reagent [3,4].

The choice of cut off value for blood specific IgE has not been mentioned in guidelines, so Serum specific IgE is often overused and misused for diagnostic allergy. The cut off value for sIgE that distinguishes between an allergic and non-allergic individual has been difficult to define. Serum specific IgE antibody to different allergens have different cut off values depending on the type of the allergens, nature of allergens, concentration of allergen exposure and duration of allergens exposure. Children raised with high exposure to dust mite allergens will have high titer of sIgE antibodies to dust mite allergens [5,6].

There is a strong association between acute episodes of respiratory allergy (Allergic Asthma) with increase in total IgE > 250 IU/MI). Most individuals have very low level of circulatory serum IgE (<100 IU/ml where 1 IU = 2.4 ng IgE). Allergen specific IgE should be considered as a marker for atopy and IgE sensitization but it cannot predict clinical severity of reactions and will have to be interpreted on the basis of the individual’s case history [7,8].

There is a complex interaction of neuroendocrine and metabolic immune system. Inhaled corticosteroids are the mainstay treatment for respiratory allergy and there is an increasing concern about their systemic side effects especially adrenal suppression. Basal serum cortisol is an indirect biomarker for detecting suppression of HPAA and the disease severity [9].

Peripheral blood eosinophilia is a hallmark of severe allergic disease and correlates with severity of the disease due to synthesis of IL5—prominent Th2—driven immune response. Infiltrating eosinophils release MBP, ECP, and EPO which in turn injure the nasal epithelial cells and induce hyper responsiveness & chronic remodeling [10,11].

Methods

Data from 75 participants coming to outpatient department of National Allergy Centre, presenting with respiratory allergy was analyzed retrospectively. The patients were without history of food allergy as recorded using a standard questionnaire.

These patients were aged between 05–70 years and comprised of 49 (65.3%) males and 26 (34.7%) females.

All these patients were also sub-categorized into different age groups (Figure 1).

The patients were without any history of food allergy as recorded using a standard questionnaire.

The diagnosis of Respiratory Allergy (allergic asthma with rhinitis) was made based on history and clinical investigations following the criteria of GINA guidelines (2014) [12,13].

Skin Prick Tests (SPT)

Data obtained from SPT tests by various allergens of these patients was recorded. Sensitization to food (7 Allergens) and common environmental allergens (35 standardized): (Pollen/Fungi/Dust mite/ Cockroach) (Allergo Pharma & Greer lab Inc.) were tested. The skin test reactions were tested with standardized allergens and were graded after 20 minutes in comparison to the wheal size of positive control, i.e. histamine diphosphate (10 mg/ml). In which maximum horizontal diameter of wheal >4 mm was considered as immunologically significant positive reaction. Why is there a low level of agreement between SPT & s pIgE in case of fungi & food allergens? It could be because of the presence of different extracts of proteins and Epitopes present in the commercial SPT kits. In our study SPT was done with Alk Lancet which is reliable, tolerable and comparable with other devices [14,15].

Total IgE estimation: Serum total IgE estimation was carried out for each patient by ImmunoCAP® in 75 subjects

Specific IgE estimation: Sera of patients showing marked positive skin reactions were evaluated by ImmunoCAP® for specific IgE against seven food allergens and nine aeroallergens.

Statistical analysis

The statistical analysis was performed using SPSS (Statistical Package for Social Sciences) version 10.5 software. The data was analyzed for all variables using the Chi-square test.

Comparison evaluation

1. Sensitization pattern based on Serum (16 specific IgE >1 kU/ml) with Skin prick tests Positive >4 mm Wheal Size (35 aero-allergens).

Figure 1: Demography – Age Distribution.
2. Serum total IgE levels >250 IU/ml with Sensitizations based on Serum (16 specific IgE >1 kU/ml) with Skin prick tests Positive >4 mm Wheal Size (35 aero-allergens)

3. Absolute eosinophil count >200 cells/µL with Sensitizations based on Serum (16 specific IgE >1 kU/ml) with Skin prick tests Positive >4 mm Wheal Size (35 aero-allergens)

4. Serum cortisol level <10 mcg/dL with Sensitizations based on Serum (16 specific IgE >1 kU/ml) with Skin prick tests Positive >4 mm Wheal Size (35 aero-allergens).

Results

1. Sensitization pattern based on Serum (16 specific IgE >1 kU/ml) with Skin prick tests Positive >4 mm Wheal Size (35 aero-allergens).

75 subjects were analyzed for respiratory allergy against foods & Aero-allergens in which poly-sensitization was seen in maximum patients (n=60, 80%) (Figure 2), we further did analysis for allergen categorization in these patients.

Data observed for Monosensitized and Poly-sensitized patterns demonstrated that maximum allergic cases were of dust mites (n=10, 66.6%) & both mites and cockroach (n=57, 95%) respectively (Figure 3).

The combined results for both tests (SPT>4 mm and sIgE >1 kU/ml) demonstrated a significantly higher sensitivity. Data below showed a high level of agreement between SPT and sIgE Figures 4-8.
sensitization in physician based diagnosis of respiratory allergy which differs from the International threshold cut off criteria of SPT > 3 mm based on nasal provocation test [4,21].

In the current study of 75 subjects of respiratory allergy both SPT & sIgE were carried out to detect the level of agreement (Concordance Vs. Discordance), and their relevance with Total IgE, AEC and S. Cortisol.

Sensitization was considered immunologically significant, if SPT showed >4 mm wheal size & sIgE was >1 kU/ml and a higher percentage of sensitivity and PPV was observed (Average Percentage: Sensitivity – 89.3%, PPV – 69.28%) on combining both the tests.

House Dust Mite (D. pteronyssinus, D. farinae) (n=67, 89.33%) was observed to have high sensitivity 100% & Positive predictive value (PPV) 74.3% which displayed a high level of agreement between SPT & sIgE.

Cockroach (n=57, 76%) also showed a high level of agreement between SPT & sIgE with sensitivity up to 100% & PPV as 80.31%.

Pollens (n=56, 74.6%), showed a high level of agreement between SPT & sIgE with sensitivity as high as 91.2% & PPV as 83.3%.

Fungi (n=21, 28%), showed low level of agreement between SPT & sIgE low level of sensitivity 76.7% & low level of PPV 49.7%, with a high NPV Negative Predictive Value 55.71%.

In case of food allergens there was no agreement between SPT & sIgE with high specificity of 100% & Negative predictive value (NPV) of 100% for detection of respiratory allergy.

In our study both SPT >4 mm sIgE >1 kU/ml have high level of agreement between House Dust Mite, Cockroach & Pollens, but low level of agreement between fungi & food allergens. Our results verify that cut off value of wheal size of >3 mm & sp IgE > 0.35 kU/ml cannot explain the clinical sensitivity of patients from respiratory allergy.

Discussion on data analysis- Relevance to total IgE, AEC & S. cortisol

In our current study of 75 subjects (n=60, 80%) were poly-sensitized, among which (n=67, 89.33%) (Figure 4) were found to be significant positive with House Dust Mites (D. pteronyssinus, D. farinae) (n=41, 54.3% Sensitivity 100%, PPV 73.68%), D. farinae n=43, 54.3% sensitivity 100%, PPV 75.44%). Mixed- D. pteronyssinus + D. farinae n=67, 89.3% Normal SPT<3mm, sIgE<0.35 ku/ml [4].
farinae, D. pteronyssinus), Cockroach (n=57, 76%) (Figure 7) & Pollens (n=56, 74.6%) (Figure 6). This explains the increase in exposure to the different aero allergens suggestive of severity of the disease.

High titer >250 IU/ml of Total IgE was found in 58.6% of subjects (Figure 9). This verifies strong association between elevated S. Total IgE & Respiratory allergy (allergic rhinitis & Asthma) in Indian patients. Similar observation was found in various studies.

Increased Absolute Eosinophil count >200 cells /ul was found in 57.33% (Figure 11) among poly-sensitized subjects, which supports the prominent role of Eosinophils & maintenance of ILC2 (Innate Lymphocyte Cell Type-2) and Th2 mediated cytokines (IL-5 & IL-13).

Low level of S. Cortisol <10 mcg/dL (N=22, 29.3%) (Figure 10) was found. Which may be due to the complex interaction of neuro endocrine & immune system or due to exogenous irregular intake of oral corticosteroids [21]. To the best of our knowledge there is no published data attributable to find the significance of lower cortisol level in polysensitized respiratory allergic patients.

Conclusion

Our study has several strengths and limitations that are worth acknowledging. One strength is that our study was based upon data collected from a nationally representative population with a wide range of age. This allowed us to directly compare sensitization rates in subjects of different ages suffering from respiratory allergies (allergic rhinitis bronchial asthma). Secondly, sensitization of subjects was confirmed by a standardized measurement of allergen-specific IgE and skin prick test (cutoff for sensitization considered as SPT >4 mm wheal Size & specific IgE >1 kU/ml). Thirdly, we limited our subjects to those with a history of physician diagnosed respiratory allergic diseases and with active symptoms, and excluded patients with possible non allergic diseases.

Based on the current study and the data analysis following conclusions can be drawn

a. Skin prick test and Specific IgE concordance (cutoff for sensitization considered as SPT >4 mm wheal Size & specific IgE >1 kU/ml) were found to be immunologically significant. Both tests are equally acceptable with higher sensitivity levels 100% and higher percentage of Positive Predictive Values (PPV) 74.6 %. There was high agreement between both the tests in cases of mites, pollen & cockroach and low levels of agreement in case of fungi & food allergens. Poly sensitization based on specific IgE and SPT (Figure 4) had maximum cases of dust mite (n=67, 89.33%), fungi (n=21, 28%), pollen (n=56, 74.6%), Cockroach (n=57, 76%).

b. Poly-sensitization has strong association with high levels of total IgE>250 IU/ml (n=44, 58.6%) (Figure 9), AEC level cut off >200 cell/uL (n=43; 57.33%) (Figure 11) and also low level of S. cortisol 10pg/ml (n=38; 50.6%) (Figure 10) suggestive of ongoing inflammation and remodeling with HPA Suppression.

c. When these biomarkers (Total IgE, S. cortisol, AEC) were taken for differentiating Mono allergen-sensitized from Poly allergen-sensitized, low PPV % (95%CI) were observed: 38.2 (21.9-54.6), 28.2 (14.1-42.3), 28.6 (14.9-42.2) for Total IgE, S. cortisol, and AEC respectively and percentage for correctly differentiating Mono allergen-sensitized from Poly allergen-sensitized were found to be 17.3, 14.7 and 16.0 for these biomarkers respectively.

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References


